

#### **Short Communications**

# First record of *Xestochironomus* Sublette and Wirth, 1972 (Chironomidae: Chironominae) in the Mexican Nearctic with notes on their habitat

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Academic editor: Gunnar Kvifte

Received: 03 Dec 2018 | Accepted: 19 Feb 2019 | Published: 01 Apr 2019

Citation: Bello-González O, Alonso-EguíaLis P, Mercado-Silva N (2019) First record of *Xestochironomus*Sublette and Wirth, 1972 (Chironomidae: Chironominae) in the Mexican Nearctic with notes on their habitat. Biodiversity Data Journal 7: e32124. <a href="https://doi.org/10.3897/BDJ.7.e32124">https://doi.org/10.3897/BDJ.7.e32124</a>

#### Abstract

We report the first record of *Xestochironomus* Sublette and Wirth, 1972 for the Mexican Nearctic. Larvae of *Xestochironomus* are known from the Neotropics and Nearctic regions. We report them for the Sonora river, NW Mexico, 300 km SW from the closest previous record in the U.S. Habitat data are provided and discussed. Our finding provides supporting evidence for the continuous presence of the genus throughout the Americas, including desert systems.

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# Introduction

The Chironomidae are widely distributed throughout the world (Ashe et al. 1987, Ferrington 2008). Chironomids are one of the most diverse and ecologically important groups found in freshwater systems. They play key roles in community and ecosystem functioning (Porinchu and MacDonald 2003).

Xestochironomus Sublette and Wirth, 1972 substituted *Insulanus* Sublette, 1967 after a study on adults mostly from the Antillean islands (Sublette and Wirth 1972). The genus includes species with larvae highly specialized on xylophagy, inhabiting lotic environments with little human disruption in the American continent (Borkent 1984). *Xestochironomus* has mainly been reported for tropical environments in the Neotropicsand in the S and SW United States. (Nearctic region) (Borkent 1984, Hudson et al. 1990, Pinho and Souza 2013). With Mexico being a transition zone between the neotropic and nearctic zoogeographic regions, presence of a member of the genus was expected. It was, however, not reported in Reiss (1982), Spies and Reiss (1996) or Spies et al. (2009), the main sources for data on Chironomidae from Mexico. The genus has only been reported once in Mexico, for the Calakmul Biosphere Reserve in the southern State of Campeche (Contreras-Ramos and Andersen 1999), >2000 km SE from our current record. This is thus the first record for *Xestochironomus* for the Mexican Nearctic.

# Methods

Collection sites are located in the Sonora and Bacanuchi rivers (Fig. 1). Samples were obtained in November 2017 with a D-net (40 cm wide; 0.5 mm mesh) and via vigorous washing of woody debris hand-picked from the collection site. Samples were preserved in 80% ethanol. In the laboratory, individuals were separated from the debris and mounted on microscope slides in Euparal following Saether (1969). Slides were then examined under an optical microscope (Zeiss, model: Primo Star) with Nomarsky phase contrast and 1000x magnification with an immersion oil objective, coupled to a AxioCam ERc 5s camera. Borkent (1984), Epler (2001) and Ferrington et al. (2008) were used for specimen identification.

Habitat variables were obtained during the field collections. Bottom substrate was classified using Wentworth's scale (Cummins 1996). Temperature (°C), pH, dissolved oxygen (mg/l) and conductivity (units) were obtained with a YSI Professional Plus (Xylem Inc) multimeter. Water depth and velocity at the collection site were measured with a Flowmate 2000 (Marsh-McBirney Inc) flowmeter.

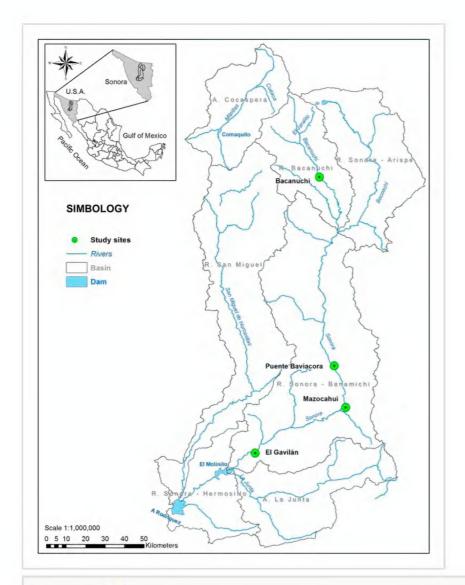


Figure 1. doi

Localities where *Xestochironomus* larvae were captured in the Sonora River, north-western Mexico.

# Xestochironomus sp.

#### Materials examined

Mexico: Sonora: Bacanuchi (30°35'57"N, -110°14'36"W), collectors: P. Alonso-EguíaLis and O. Bello, 9 individuals. Deposited at the scientific collection of the Laboratorio de Bioindicadores of the Instituto Mexicano de Tecnología del Agua. Captured 18 November 2017.

México: Sonora: Puente Baviacora (29°43'32"N, -110°10'30"W), collectors: P. Alonso-EguíaLis and O. Bello, 3 individuals. Deposited at the scientific collection of the Laboratorio de Bioindicadores of the Instituto Mexicano de Tecnología del Agua. Captured 12 November 2017.

México: Sonora: Mazocahuí (29°31'58"N, -110°07'17"W), collectors: P. Alonso-EguíaLis and O. Bello, 1 individual. Deposited at the scientific collection of the Laboratorio de Bioindicadores of the Instituto Mexicano de Tecnología del Agua. Captured 12 November 2017.

México: Sonora: El Gavilán (29°19'18"N, -110°32'25"W), collectors: P. Alonso-EguíaLis and O. Bello, 1 individual. Deposited at the scientific collection of the Laboratorio de Bioindicadores of the Instituto Mexicano de Tecnología del Agua. Captured 17 November 2017.

#### Identification

Xestochironomus larvae are similar to those of Stenochironomus Kieffer, 1919. However, they can be distinguished by the following attributes (all of which were present in the individuals used for this article): sclerotised mentum concave with well sclerotised teeth (Fig. 2a); mentoventral plates vestigial; anal tubules elongated with 4-5 constrictions (Fig. 2b); cephalic capsule dorsoventrally flattened with a Y-shaped dorsal design (Fig. 2c) and antennal blade extending beyond the apex of the third antennal segment (Fig. 2d). In contrast, Stenochironomus larvae have 10-12 teeth in the mentum, the antennal blade reaches only the apex of the 2nd antennal segment and the anal tubules have, at most, two constrictions.

#### **Environmental variables**

Habitat variables for collection sites are presented in Table 1.

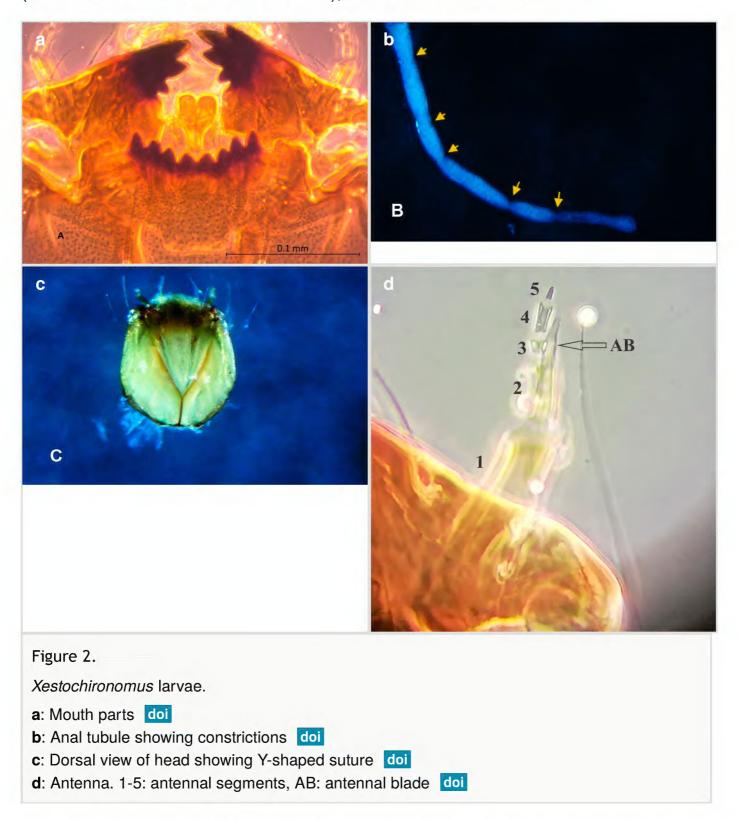
Table 1.
Habitat data for sites where Xestochironomus larvae were found. Site = SIS; Bacanuchi = BC;
Puente Baviacora = PB; Mazocahui = MZ; El Gavilán = EG; Conductivity = Cond; Dissolved oxygen
= DO; temperature = T; depth = D; water velocity = V; substrate = S; sand = SN; gravel = GV.

SIS	Altitude (m)	Cond (µS/cm)	рН	DO (mg/l)	T (° C)	D (cm)	V (m/s)	s
вс	1030	784	7.4	6.22	23.4	10	0.15	SN-GV
РВ	552	860	7.1	4.26	24.2	8	0.44	SN
MZ	473	1249	7.9	3.48	30.9	5	0.20	SN
EG	328	1508	7.8	3.23	27.2	9	0.16	SN

### Discussion

Members of *Xestochironomus* are known from the Americas with most records from neotropical areas in Brazil, Chile, Colombia, Costa Rica, Cuba, Dominica, Guatemala, Jamaica, Panama, Peru, Puerto Rico and Venezuela. Nearctic records are all from the USA in Florida, South Carolina, Georgia, Texas and New México (Andersen and Kristoffersen 1998, Bello-González et al. 2016, Borkent 1984, Hudson et al. 1990, Pinho and Souza 2013, Ruiz-Moreno et al. 2000, Sublette and Sasa 1994, Sublette and Wirth 1972). The nearest record to the one presented here is from approximately 300 km away in the San Francisco Hot Springs area of the Gila River in New Mexico (Borkent 1984). In

Mexico, the only record is for the Calakmul biosphere reserve in the Yucatan Peninsula (Contreras-Ramos and Andersen 1999), over 2000 km SE of our record.



Following Abell et al. (2008) ecoregion classification, most *Xestochironomus* records are from humid tropical or subtropical areas. Some of the species in this genus can be very abundant in tropical streams draining rainforests (Ferrington et al. 1993, Grund 2006). These conditions favour woody debris inputs to the channel, which constitute both habitat and a food resource for larvae of *Xestochironomus* (Epler 2001, Sanseverino and Nessimian 2008). High densities in larval xylophagous chironomids are found under such conditions (Cranston 2008).

The Sonora River basin, located in the Gulf of California climatic province, has a distinctive dry climate (Vidal 2005). Vegetation types dominating this area include shrubs and herbs

(Martínez-Yrízar et al. 2009) with Cottonwood (*Populus* sp.) being an important component of the riparian vegetation. These dry conditions lead to relatively low woody debris inputs to stream channels (Bunn et al. 2006, Davies et al. 1995, Jones 1997, Cushing and Allan 2001). Our record from the Sonora and records from the Gila system nevertheless confirm that larvae of *Xestochironomus* can occur in streams located in arid regions with little input of woody debris. It is thus possible that the genus has a continuous distribution from the Neotropics to the Nearctic region, with deserts in N Mexico and SW USA not being barriers to their distribution.

Most (12/14) captured larvae were collected from sites Bacanuchi and Puente Baviacora, while only one individual was captured in sites Mazocahui and El Gavilan despite all sites having similar sampling efforts. Other than the information related to trophic habits, very little data exists about the overall conditions of the habitat required by *Xestochironomus* larvae. Since they typically inhabit galleries in submerged wood, their low mobility could render them sensitive to changes in local conditions. Xylophagous larvae usually require good water quality (Borkent 1984, Cranston 2008). Mazocahui and El Gavilan had relatively high conductivity (>1000  $\mu$ S/cm), values above that threshold are typically associated with pollution (Chapman and Kimstach 1996). Low dissolved oxygen and high temperatures in these sites might also have resulted in little oxygen being available for aquatic fauna. Thus, lower water quality in Mazocahui and El Gavilan might be a cause for *Xestochironomus* having a relatively lower abundance in these sites.

# Acknowledgements

The authors would like to thank Buenavista del Cobre S.A. de C.V. for field work support. V. Ramírez, F. Aviles, S. González, R. Ojeda and F. Bravo assisted with field and laboratory work. T. Andersen assisted with bibliographic work. We thank Dr. Gunnar Kvifte, Dr. Trond Andersen and especially Dr. Viktor Baranov for his comments on our work. This manuscript is a result of OCBG's Master's degree at CIByC – UAEM, co-advised by NMS and PEAE.

# **Author contributions**

OCBG. Field work, sample processing and ID, manuscript preparation.

PEAE. Field work, manuscript preparation.

NMS. Field work, manuscript preparation.

# References

Abell R, Thieme ML, Revenga C, Bryer M, Kottelat M, Bogutskaya N, Coad B, Mandrak N, Balderas SC, Bussing W, Stiassny MLJ, Skelton P, Allen GR, Unmack P, Naseka A, Ng R, Sindorf N, Robertson J, Armijo E, Higgins JV, Heibel TJ, Wikramanayake E, Olson D, López HL, Reis RE, Lundberg JG, Sabaj Pérez M, Petry P (2008) Freshwater

- ecoregions of the world: A new map of biogeographic units for freshwater biodiversity conservation. BioScience 58 (5): 403-414. <a href="https://doi.org/10.1641/b580507">https://doi.org/10.1641/b580507</a>
- Andersen T, Kristoffersen L (1998) New species of Xestochironomus Sublette and Wirth (Chironomidae: Chironominae) from Chile and Costa Rica. Journal of the Kansas Entomological Society 71 (3): 296-303.
- Ashe P, Murray DA, Reiss F (1987) The zoogeographical distribution of Chironomidae (Insecta: Diptera). Annales de Limnologie 23 (1): 27-60. <a href="https://doi.org/10.1051/limn/1987002">https://doi.org/10.1051/limn/1987002</a>
- Bello-González OC, Andersen T, Hagenlund LK (2016) A new species of Xestochironomus Sublette and Wirth, 1972 from Cuba (Diptera, Chironomidae). Norwegian Journal of Entomology 63: 44-49.
- Borkent A (1984) The systematics and phylogeny of the Stenochironomus complex ( Xestochironomus, Harrisius, and Stenochironomus) (Diptera: Chironomidae). Memoirs of the Entomological Society of Canada 128: 1-269.
- Bunn S, Balcombe S, Davies P, Fellows SC, McKenzie-Smith JF (2006) Aquatic productivity and food webs of desert river ecosystems. In: Kingsford R (Ed.) Ecology of Desert Rivers. Cambridge University Press, UK, 76-99 pp.
- Chapman D, Kimstach V (1996) Selection of water quality variables. In: Chapman D (Ed.) Water Quality Assessments A Guide to Use of Biota, Sediments and Water in Environmental Monitoring. Second Edition. World Health Organization. UNESCO. University Press Cambridge, 74-133 pp.
- Contreras-Ramos A, Andersen T (1999) A survey of the Chironomidae (Diptera) of Kalacmul Biosphere Reserve, Mexico. Chironomus. Newsletter of Chironomid Research 12: 3-5.
- Cranston PS (2008) Phylogeny predicts larval biology in a wood-mining chironomid (Diptera: Chironomidae). African Entomology 16 (1): 1-6. <a href="https://doi.org/10.4001/1021-3589-16.1.1">https://doi.org/10.4001/1021-3589-16.1.1</a>
- Cummins KW (1996) Invertebrates. In: Petts G, Calow P (Eds) River Biota. Diversity and Dynamics. Blackwell Science, Oxford, 75-91 pp.
- Cushing CE, Allan JD (2001) Desert rivers of the Southwest. Streams: Their Ecology and Life. Academic Press, San Diego, California, 128-139 pp.
- Davies BR, O'Keefe JH, Snaddon CD (1995) River and stream ecosystems in southern Africa: predictably unpredictable. In: Cushing CE, Cummins KW (Eds) River and Stream Ecosystems. Elsevier Science B.V, The Netherlands, 537-600 pp.
- Epler JH (2001) Identification Manual for the Larval Chironomidae (Diptera) of North and South Carolina. A Guide to the Taxonomy of the Midges of the Southeastern United States, Including Florida. Special Publication SJ2001-SP13. North Carolina Department of Environment and Natural Resources, Raleigh, NC, and St. Johns River Water Management District, Palatka, Florida, 526 pp.
- Ferrington L, Buzby K, Masteller E (1993) Composition and temporal abundance of Chironomidae emergence from a Tropical Rainforest Stream at El Verde, Puerto Rico. Journal of the Kansas Entomological Society 66 (2): 167-180.
- Ferrington L (2008) Global diversity of non-biting midges (Chironomidae; Insecta-Diptera) in freshwater. Hydrobiologia 595 (1): 447-455. <a href="https://doi.org/10.1007/s10750-007-9130-1">https://doi.org/10.1007/s10750-007-9130-1</a>

- Ferrington LC, Berg MB, Coffman WP (2008) Chironomidae. In: Merritt RW, Cummins KW, Berg MB (Eds) An Introduction to the Aquatic Insects of North America. 4th Edition. Kendall/Hunt Publishing Company, Dubuque, 847-989 pp.
- Grund M (2006) Chironomidae (Diptera) in Dominican amber as indicators for ecosystem stability in the Caribbean. Palaeogeography, Palaeoclimatology, Palaeoecology 241: 410-416. <a href="https://doi.org/10.1016/j.palaeo.2006.04.005">https://doi.org/10.1016/j.palaeo.2006.04.005</a>
- Hudson PL, Lenat DR, Caldwell BA, Smith D (1990) Chironomidae of the Southeastern United States: A checklist of species and notes on biology, distribution, and habitat. Fish and Wildlife Research 7: 1-46.
- Jones JB (1997) Benthic organic matter storage in streams: Influence of detrital import and export, retention mechanisms, and climate. Journal of the North American Benthological Society 16 (1): 109-119. <a href="https://doi.org/10.2307/1468243">https://doi.org/10.2307/1468243</a>
- Martínez-Yrízar A, Stephen RF, Búrquez A (2009) Los ecosistemas terrestres de Sonora: un diverso capital natural. In: Molina-Freaner F, Van-Devender T (Eds)
   Diversidad biológica del estado de Sonora. UNAM, Mexico, 129-156 pp.
- Pinho LC, Souza JF (2013) New species of Xestochironomus Sublette et Wirth, 1972 (Diptera: Chironomidae) from Atlantic Forest, Brazil. Zootaxa 3652 (5): 595. <a href="https://doi.org/10.11646/zootaxa.3652.5.9">https://doi.org/10.11646/zootaxa.3652.5.9</a>
- Porinchu D, MacDonald GM (2003) The use and application of freshwater midges (Chironomidae: Insecta: Diptera) in geographical research. Progress in Physical Geography 27 (3): 378-422. <a href="https://doi.org/10.1191/030913303767888491">https://doi.org/10.1191/030913303767888491</a>
- Reiss F (1982) Chironomidae. In: Hurlbert S, Villalobos-Figueroa A (Eds) Aquatic biota of Mexico, Central America and the West Indies. San Diego State University, California, 433-438 pp.
- Ruiz-Moreno J, Ospina-Torres R, Riss W (2000) Guía para la identificación genérica de larvas de quironómidos (Diptera-Chironomidae) de la Sabana de Bogotá. II. Caldasia 22 (1): 15-33.
- Saether OA (1969) Some Nearctic Podonominae, Diamesinae and Orthocladiinae (Diptera: Chironomidae). Bulletin of the Fisheries Research Board of Canada 170: 1-154.
- Sanseverino A, Nessimian J (2008) The food of larval Chironomidae (Insecta, Diptera) in submerged litter in a forest stream of the Atlantic Forest (Rio de Janeiro, Brazil). Acta Limnologica Brasiliensia 20 (1): 15-20.
- Spies M, Reiss F (1996) Catalog and bibliography of Neotropical and Mexican Chironomidae (Insecta, Diptera). Spixiana (Supplement) 22: 61-119.
- Spies M, Andersen T, Epler JH, Watson CN (2009) Chironomidae (non-biting midges).
  In: Brown BV, Borkent A, Cumming JM, Wood DM, Woodley NE, Zumbado M (Eds)
  Manual of Central American Diptera . 1. NRC Research Press, Ottawa, Ontario,
  Canada, 437-480 pp.
- Sublette JE, Wirth WW (1972) New genera and species of West Indian Chironomidae (Diptera). The Florida Entomologist 55 (1): 1-17. <a href="https://doi.org/10.2307/3493636">https://doi.org/10.2307/3493636</a>
- Sublette JE, Sasa M (1994) Chironomidae collected in onchocerciasis endemic areas of Guatemala (Insecta, Diptera). Spixiana Supplement 20: 1-60.
- Vidal R (2005) Las regiones climáticas de México. Temas selectos de geografía de México. Instituto de Geografía-UNAM, México, 210 pp.